Hematoxylin and Eosin (H+E) Staining

I. paraffin sections must be deparaffinized and rehydrated
   **(xylene, iso-propanol, 96%,75%,50%alcohol, a.dest.;
   5 min each)
II. cryo sections must be fixed

Nuclei Staining (Hematoxylin Staining)

Also used for counter staining after AEC or DAB Chromophore

1. 30 sec.-2 min Hematoxylin
2. 1 min demineralised water) –slightly agitating
3. 30 sec.HCl- alcohol (150ml dem. Water+~ 30ml HCl-
   alcohol) –slightly agitating
4. 30 sec dem. Water –slightly agitating
5. 1-2 min. fresh tape water ) –slightly agitating
6. 30 sec. dem. Water

Slides can now be covered. Hematoxylin is insoluble in aqueous mounting media.
If organic mounting media is to be used, slides must be dehydrated. **(see above,
in opposite direction)
Over staining must be avoided. If staining is too weak, repeat the above.)

Cytoplasm (Eosin Staining)

1. 20-30 sec. Eosin) –slightly agitating
2. 5-10 sec. 50% alcohol –slightly agitating
3. 5-10 sec. 75% alcohol –slightly agitating
4. 10-20 sec. 96% alcohol –slightly agitating
5. 20-30 sec. iso propanol ) –slightly agitating, tap off access propanol,
   (microscopic control is recommended)
6. 30-1 min sec. xylene
7. 30 sec. xylene

Slides now must be mounted in organic mounting media such as Entelan, eosin is
highly soluble in water. Over staining can be removed by rinsing in water or 50% alcohol.